

Cancer Research

OncomiR or Tumor Suppressor? The Duplicity of MicroRNAs in Cancer

Alexander A. Svoronos¹, Donald M. Engelman¹, and Frank J. Slack²

Abstract

MicroRNAs (miRNA) are short, noncoding RNAs whose dysregulation has been implicated in most, if not all, cancers. They regulate gene expression by suppressing mRNA translation and reducing mRNA stability. To this end, there is a great deal of interest in modifying miRNA expression levels for the treatment of cancer. However, the literature is fraught with inconsistent accounts as to whether various miRNAs are oncogenic or tumor suppressive. In this review, we directly examine these inconsistencies and propose several mechanisms to explain them. These mechanisms include the possibility that specific miRNAs can simultaneously produce

Introduction

MicroRNAs (miRNA) are short, 18-25 nucleotide-long, noncoding RNA molecules that regulate gene expression by suppressing mRNA translation and reducing mRNA stability, usually through imperfect complementary base pairing to the 3'-untranslated region. Since their 1993 discovery in C. elegans, it has become ever more apparent that miRNAs are dysregulated in most, if not all, cancers. Many of these miRNAs either contribute to or repress the cancer phenotype by inhibiting the expression of tumor suppressors or oncogenes, respectively. Generally, oncogenic miRNAs (oncomiRs) are overexpressed in cancers while tumor-suppressive miRNAs are underexpressed. When these oncomiRs or tumor-suppressor miRNAs are inhibited or stimulated, respectively, cancer cell proliferation, metastasis, and/or survival may be significantly reduced, depending on the type of cancer and the specific miRNA being affected. It is even possible for cancers to become completely reliant upon, or "addicted", to an oncomiR, such that suppression of the oncomiR results in complete regression of the cancer (1). Thus, miRNAs have classically been categorized as either oncogenic or tumor suppressive, and controlling their expression for therapeutic purposes is the subject of intense ongoing research.

However, there is reason to propose that the therapeutic approaches should proceed with caution. The literature is fraught

©2016 American Association for Cancer Research.

competing oncogenic and tumor suppressive effects by suppressing both tumor suppressive mRNAs and oncogenic mRNAs, respectively. In addition, miRNAs can modulate tumor-modifying extrinsic factors, such as cancer-immune system interactions, stromal cell interactions, oncoviruses, and sensitivity to therapy. Ultimately, it is the balance between these processes that determines whether a specific miRNA produces a net oncogenic or net tumor suppressive effect. A solid understanding of this phenomenon will likely prove valuable in evaluating miRNA targets for cancer therapy. *Cancer Res; 76*(13); 3666–70. ©2016 AACR.

with conflicting reports as to whether specific miRNAs are oncogenic or tumor suppressive. Repeatedly, certain miRNAs have been shown to be oncogenic in one scenario, but tumor suppressive in another. Diversity of effects is not a surprise given the large number of genes influenced by a particular miRNA. It hence follows that the classification of a miRNA as oncogenic or tumor suppressive may represent an oversimplification that must be carefully scrutinized in all cancer miRNA studies. To date, this issue has received little consideration, and few studies have directly examined its potential causes.

Here, we highlight several examples in which a specific miRNA can act either as a tumor suppressor or an oncogene, depending on the context. We attempt to explain the phenomenon via examination of the affected cellular and molecular mechanisms, and we propose multiple factors that can influence whether a miRNA has a net oncogenic or net tumor suppressive effect. Finally, we argue for a holistic approach in examining the effects of miRNAs in cancer that incorporates the interactions of the miRNA's multiple targets and effects beyond the tumor cell, including interactions with the immune system, stromal cells, and therapy.

A Net Effect: The Targeting of Both Tumor Suppressors and Oncogenes

As an example of a miRNA that can act as either an oncomiR or a tumor suppressor depending on the context, we consider miR-125b. miR-125b acts as an oncomiR in the vast majority of hematologic malignancies but as a tumor suppressor in many solid tumors (2, 3). This apparent paradox can be reconciled by taking into account the fact that a single miRNA molecule has the capacity to target tens to hundreds of different mRNAs, some of which may have opposing oncogenic or tumor-suppressive functions. In the case of miR-125b, targets include mRNAs encoding antiapoptotic factors (MCL1, BCL2L2, and BCL2), proapoptotic factors (TP53, BAK1, BMF, BBC3, and MAPK14), proproliferative factors (JUN, STAT3, E2F3, IL6R, and ERBB2/3), metastasis promoters (MMP13, LIN28B, and ARID3B), metastasis inhibitors



¹Department of Molecular Biophysics and Biochemistry, Yale University, New Haven, Connecticut. ²Institute for RNA Medicine, Departments of Pathology and Medicine, Beth Israel Deaconess Medical Center/Harvard Medical School, Boston, Massachusetts.

Corresponding Authors: Frank J. Slack, Beth Israel Deaconess Medical Center 3 Blackfan Circle, CLS412 Boston, MA 02215. Phone: 617-735-2601; Fax: 617-735-2646; E-mail: fslack@bidmc.harvard.edu; and Donald M. Engelman, donald.engelman@vale.edu

doi: 10.1158/0008-5472.CAN-16-0359

(STARD13, TP53INP1, and TP53), and factors involved in hematopoietic differentiation (CBFB, PRDM1, IRF4, IL2RB, and IL10RA; refs. 2, 3). Hence, it is likely that it is the balance of expression of these oncogenes/tumor suppressors that determines whether miR-125b will have a net oncogenic or net tumorsuppressive effect within an individual cancer. It is plausible that miR-125b largely exerts its oncogenic role in hematopoietic malignancies via the suppression of hematopoietic differentiation factors, which are of limited importance in most solid tumors. Combined with the suppression of miR-125b's other tumor-suppressive targets, the tumor-suppressive effects of miR-125b are trumped to produce a net oncogenic effect. For solid tumors, whether miR-125b is oncogenic or tumor suppressive is far more variable, likely because there is a more even balance between its oncogenic and tumor suppressive effects. Interestingly, the strong oncogenic role of miR-125b in hematologic malignancies can be overcome in certain instances. One notable example is chronic lymphocytic leukemia (CLL), in which miR-125b has been demonstrated to have a tumor-suppressive role (4). Although the possibility has not been directly investigated, one plausible explanation for this anomaly is that high overexpression of BCL2, an antiapoptotic target of miR-125b, is a critical hallmark of CLL (5). Thus, its suppression by miR-125b may be enough to tip the scales so that a net tumor-suppressive effect is produced.

Another excellent example of this phenomenon is provided by miR-155. By and large, miR-155 is considered an oncomiR. It possesses an oncogenic role in a large number of solid and hematologic malignancies (6-8), and its overexpression alone in lymphoid tissues is sufficient to produce an aggressive disseminated lymphoma in a miR-155 Cre-loxP tetracycline-controlled knock-in mouse model (6, 7). Consistent with miR-155's oncogenic potency, the lymphoma produced in these mice was demonstrated to be "addicted" to, or completely reliant upon, miR-155; tetracycline-induced withdrawal of miR-155 resulted in complete regression of the lymphoma (6). Nevertheless, despite its strong oncogenic effects, there is evidence that miR-155 has a tumor-suppressive role in some cancers. Levati and colleagues demonstrated that miR-155 inhibits proliferation in several melanoma cell lines, due in part to suppression of SKI, a commonly overexpressed oncogene in melanoma (9). Similarly, Li and colleagues and Qin and colleagues demonstrated that miR-155 exerts a tumor-suppressive effect in gastric cancer and ovarian cancer-initiating cells via targeting of SMAD2 and CLDN1, respectively (10, 11). In addition, Palma and colleagues found that miR-155 has a proapoptotic and prodifferentiation role in FLT3wildtype normal karvotype acute myeloid leukemia (NK-AML), which confirms that miR-155's tumor-suppressive role is not confined to solid malignancies (12). Strikingly, this was not the case for NK-AML harboring the FLT3-ITD mutation, which exhibited overexpression of miR-155 (12).

This set of observations highlights the variability of whether a miRNA is oncogenic or tumor suppressive, even within a single cancer type. Such variability is especially apparent when one considers that sequence variations are possible within the miRNA target sites of regulated genes. Multiple studies have shown that mutations and single-nucleotide polymorphisms can result in the functional loss of existing miRNA target sites (13, 14), as well as the creation of new miRNA target sites (15). Given the genetic heterogeneity of many tumors, it is hypothetically possible for a miRNA to exert opposing effects in different regions of an individual tumor, although examples of this have yet to be observed.

miRNA Interactions with Tumor-Modifying Extrinsic Factors: The Necessity of a Holistic Approach

The growth and spread of a cancer is not solely a function of the cancer cells themselves, but also of various extrinsic factors, which interact with the cancer cells to affect their behavior. Such factors include the immune system, tumor stromal cells, therapy, and oncoviruses. MiRNAs may have large influences on each of these factors, and it is hence necessary to maintain a holistic view when examining miRNAs from a therapeutic standpoint. This view should take into account all the aforementioned factors, not just the effects of the miRNA that are specific to the cancer cells themselves. Here, we present several examples of when a miRNA exerts an oncogenic effect on a tumor-modifying extrinsic factor but a tumor-suppressive effect on the cancer cells themselves, or vice versa (summarized in Fig. 1).

Contrasting effects on tumor-immune system interactions

miRNAs may modulate the interactions of cancer cells with the immune system. It is now well established that the interaction of immune cells with cancer cells in the tumor microenvironment can play an integral role in the growth and spread of the cancer. Both Zonari and colleagues and Yu and colleagues demonstrated that miR-155 deficiency in tumor-associated macrophages promotes conversion of the macrophages from the proinflammatory, antitumoral M1 phenotype to the antiinflammatory, protumoral M2 phenotype (16, 17). Zonari and colleagues found that stable knockdown of miR-155 in the myeloid compartment of a mammary cancer mouse model resulted in increased tumor growth, and concluded that this was due to a decreased antitumoral immune response from tumor-associated macrophages (17). Similarly, Yu and colleagues observed that bone marrow transplantation from miR-155^{-/-} mice to wild-type mice resulted in increased metastasis to the lung. They attributed this to M2 tumor-associated macrophage promotion of invasion and metastasis, as evidenced by an in vitro Transwell migration assay (16). In addition, an analogous role for miR-155 was found for tumorassociated dendritic cells in ovarian cancer. Cubillos-Ruiz and colleagues selectively delivered miR-155 precursor-containing nanoparticles to ovarian cancer-associated dendritic cells by taking advantage of the dendritic cells' spontaneous enhanced endocytic activity (18). This resulted in a transformation of the dendritic cells from an immunosuppressive phenotype to an immunostimulatory phenotype that triggered a potent antitumor immune response (18). Together, these studies clearly demonstrate the importance of considering miR-155's role in tumor-infiltrating immune cells when designing miR-155based therapeutics. In the case where miR-155 suppression has a tumor-suppressive effect within the cancer cells themselves, it is likely that anti-miR-155 therapy would be optimized by targeting just the cancer cells and not the immune cells.

Two other examples of miRNAs that may have contrasting roles on tumor–immune system interactions and the cancer cells themselves are miR-30b and miR-30d. Both miRNAs target multiple oncogenes and are classified as tumor suppressors in the majority of studies that do not involve immune cells or

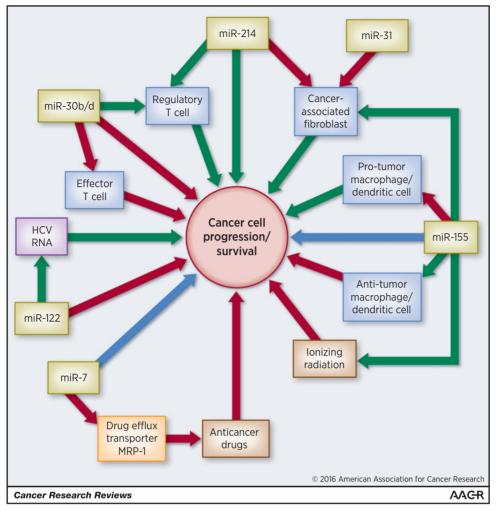


Figure 1.

Examples of miRNAs that may exert contrasting oncogenic/ tumor-suppressive effects on tumor-modifying extrinsic factors and the cancer cells themselves. Green arrows, positive regulation; red arrows, negative regulation; blue arrows either positive or negative regulation, depending on cancer cell type/context. An arrow extending directly from a miRNA to the central cancer cell refers to promotion/inhibition of cancer cell progression/survival through the miRNA's direct regulation of cancer cellendogenous mRNAs.

immunocompetent mouse models (19, 20). However, Gaziel-Sovran and colleagues found that miR-30b/d promote secretion of the immunosuppressive cytokine IL10 by directly targeting the GalNAc transferase *GALNT7* (21). The increased IL10 from miR-30b/d-mediated *GALNT7* suppression resulted in reduced T-cell recruitment and enhanced regulatory T-cell induction within tumors in an immunocompetent mouse model for melanoma (21). This reduction in antitumor immunity was accompanied by a significantly increased number of metastases (21). Thus, examining a miRNA's effect on tumor–immune system interactions may be of the utmost importance when developing a miRNA-modulating therapy for cancer.

Contrasting effects on tumor stromal cells

miRNAs may also play a role in the interaction between cancer cells and non-immune cells of the tumor stroma, such as cancerassociated fibroblasts (CAF). It is now well established that nonimmune cells of the tumor stroma can have a significant role in cancer progression. As a tumor develops, interactions between the cancer cells and the stromal cells may result in transformation of the stromal cells to a protumor state, which significantly enhances cancer growth, invasion, and metastasis. Although the mechanisms behind this transformation are poorly understood, they are partially elucidated by the discovery that miR-214 and miR-31 are

consistently downregulated, while miR-155 is consistently upregulated, in ovarian cancer CAFs compared with normal fibroblasts (22). By triple-transfecting normal fibroblasts with miR-214 and miR-31 inhibitors and pre-miR-155, Mitra and colleagues were able to convert the fibroblasts to a CAF phenotype, which, like actual CAFs, significantly increased colony formation, migration, and invasion when cocultured with ovarian cancer cells in vitro and in vivo tumor growth when coinjected with ovarian cancer cells into mice. The reverse was also true; CAFs triple-transfected with miR-155 inhibitor, pre-miR-214, and pre-miR-31 exhibited a normal fibroblast phenotype (22). Notably, the cancer-promoting effects of the CAFs and the anti-miR-214/anti-miR-31/pre-miR-155 tripletransfected normal fibroblasts were largely mediated by increased expression of the cytokine CCL5, a direct target of miR-214, thereby supporting the notion that miR-214 acts as a potent tumor suppressor through the interactions of CAFs with ovarian cancer cells (22). Nevertheless, multiple studies attribute an oncogenic role to miR-214 (23, 24). Notably, one of these studies suggested miR-214 to be an oncomiR in ovarian cancer, the same type of cancer as in Mitra and colleagues' study, through the direct suppression of TP53 (23). In addition, another of these studies demonstrated that miR-214 secreted within microvesicles by cancer cells can enter the bloodstream and exert an oncogenic effect through the induction of regulatory T cells, thereby suppressing the antitumor immune

response (24). Together, these results highlight the great complexity of this miRNA's role in cancer.

Location dependence of miRNA effects

Remarkably, the location of cancer cells can also govern whether a miRNA produces a net oncogenic or net tumor suppressive effect. This was demonstrated to be the case with miR-155 for 4T1 breast tumor cells injected into mice. Xiang and colleagues found that 4T1 cells virally transduced to overexpress miR-155 metastasized to a far lesser extent than control 4T1 cells after inoculation in the mammary fat pads of mice. Other factors, including tumor growth, remained the same. However, when the cells were injected directly into the blood stream, the miR-155-overexpressing cells produced far more metastatic lung tumors and increased tumor growth in the lungs compared with the control cells. Further analysis demonstrated that miR-155 inhibited epithelial-to-mesenchymal transition (EMT) in the cells by targeting TCF4, a key regulator of EMT. Thus, metastasis from the mammary fat pads was suppressed. However, at the same time, seeding and growth into the lung from cells that were already circulating in the bloodstream was increased, likely because the lung environment is more favorable for epithelial cell growth than it is for mesenchymal cell growth. It is possible that miR-155 overexpression promoted mesenchymal-to-epithelial transition (MET), a phenomenon many have proposed to aid the seeding of distant tissues by circulating tumor cells that have previously undergone EMT. Thus, miR-155 was tumor suppressive for 4T1 cells in the mammary fat pad but oncogenic for 4T1 cells in the blood stream and lungs. This holds important implications for the use of miRNA-modulating therapies to treat metastatic disease (25).

Contrasting effects on oncoviruses

Hepatitis C virus (HCV) infection results in the production of viral oncoproteins, chronic inflammation, and mutations in the liver that can ultimately lead to hepatocellular carcinoma. Thus, when treating a patient with hepatocellular carcinoma and concurrent HCV infection, it is important to consider a therapy's effect on not just the cancer cells, but also the HCV and immune system interactions. This may be the case with miR-122, a miRNA demonstrated to be tumor suppressive in hepatocellular carcinoma (26). Despite its antitumoral properties, miR-122 is also known to stabilize HCV RNA by binding to the 5'-untranslated region, thereby promoting HCV replication (27). In fact, Miraversen, a locked nucleic acid that inhibits miR-122 through antisense binding (27), is the most clinically advanced miRNA-targeting therapeutic in existence; it is currently in phase II clinical trials for the treatment of chronic HCV infection. Thus, treatment of hepatocellular carcinoma with exogenous miR-122 may actually enhance progression of the disease by promoting HCV replication. If a miR-122-promoting therapy is eventually developed for hepatocellular carcinoma, its use will likely require pre-screening for HCV infection. Interestingly, HCV RNA itself can be viewed as a miR-122 inhibitor by sequestering miR-122 from its host mRNA targets. Luna and colleagues demonstrated that HCV RNA functionally reduces the amount of miR-122 available for binding to its native targets, which could facilitate the oncogenic effects of HCV (28).

miRNA interactions with therapy

Interestingly, miRNAs may also affect the responsiveness to certain cancer therapies. Many studies that examine miRNA expression levels in cancer interpret a positive correlation between miRNA expression levels and increased survival as

evidence that the miRNA is tumor suppressive. However, this kind of interpretation can be misleading. For instance, the miRNA may increase the proliferation of tumor cells but at the same time confer susceptibility to a treatment, resulting in increased overall survival. This was found to be the case with miR-155 in breast cancer patients treated with ionizing radiation (29). Ionizing radiation is a therapy that works by inducing double-stranded DNA breaks in cancer cells. These doublestranded breaks can be repaired via DNA homologous recombination, and thus upregulation of enzymes involved in this mechanism can confer resistance to the therapy. Gasparini and colleagues discovered that miR-155 directly suppresses the expression of RAD51, a protein critical for DNA homologous recombination, and thereby sensitizes triple-negative breast cancer to ionizing radiotherapy (29). As a result, patients with higher miR-155 levels, despite miR-155's oncogenic effects in this cancer (8), exhibited higher overall survival (29).

In addition, multiple miRNAs have been shown to suppress drug efflux transporters, and decreases in their levels are hence associated with chemoresistance (30). While the majority of these miRNAs seem to act consistently as tumor suppressors, this is not the case for miR-7, which has been described as both a tumor suppressor (31) and an oncomiR (32) for different cancer types. However, due to its suppression of the drug efflux transporter MRP1 (multidrug-resistance associated protein 1; ref. 33), miR-7 inhibition in cancers for which it is an oncomiR could produce an overall detrimental effect, as it could enhance chemoresistance, despite slowing the growth/spread of the tumor cells. Taken together, these results strongly argue for a holistic approach when investigating the exploitation of miR-NAs as cancer therapy targets. Consideration must be given to the interactions of miRNAs with the immune system, tumor stromal cells, cancer therapies, and other factors extrinsic to the cancer cells themselves.

Conclusions

We have reviewed several of the mechanisms by which specific miRNAs can simultaneously exert competing oncogenic and tumor-suppressive effects. These effects extend from the regulation of various genes to the genes' downstream effects and to tumor-modifying extrinsic factors, such as immune system interactions and response to therapeutics. Depending on the balance between miRNA-mediated upregulation or downregulation of oncogenic and tumor-suppressive pathways, as well as the effects of the miRNA on cancer-immune system interactions and various other tumor-modifying extrinsic factors, the miRNA may produce an overall net oncogenic or net tumor-suppressive effect. A solid understanding of these mechanisms is of the utmost importance, as there is currently a great deal of excitement in the administration of exogenous miRNA mimetics and miRNA inhibitors for the control of various disease processes. This holds especially true for the field of cancer, as mounting evidence is suggesting that miRNAs are severely dysregulated in most, if not all, cancers. However, we suggest this endeavor be approached with caution, as it will likely require an excellent understanding of miRNAs from a holistic standpoint that incorporates all the aforementioned factors. To this end, we recommend the use of immunocompetent mouse models, which better replicate the tumor microenvironment, in preclinical studies of potential miRNA therapeutics. More studies investigating miRNA interactions with established

therapies would also be wise. Furthermore, due to the complexity of miRNA networks, systems biology approaches, which incorporate the interactions between various tumor-relevant systems at both the molecular and cellular scales, will be increasingly valuable. Such studies may prove necessary in order to fully utilize the vast clinical potential of miRNA therapeutics.

Disclosure of Potential Conflicts of Interest

F.J. Slack has ownership interest (including patents) in miRNA Therapeutics and Mira Dx and is a consultant/advisory board member for miRNA Therapeutics and miRagen Therapeutics. No potential conflicts of interest were disclosed by the other authors.

References

- Medina PP, Nolde M, Slack FJ. OncomiR addiction in an *in vivo* model of microRNA-21-induced pre-B-cell lymphoma. Nature 2010;467:86–90.
- Shaham L, Binder V, Gefen N, Borkhardt A, Izraeli S. MiR-125 in normal and malignant hematopoiesis. Leukemia 2012;26:2011–8.
- 3. Sun YM, Lin KY, Chen YQ. Diverse functions of miR-125 family in different cell contexts. J Hematol Oncol 2013;6:6.
- Tili E, Michaille JJ, Luo Z, Volinia S, Rassenti LZ, Kipps TJ, et al. The downregulation of miR-125b in chronic lymphocytic leukemias leads to metabolic adaptation of cells to a transformed state. Blood 2012;120:2631–8.
- Roberts AW, Seymour JF, Brown JR, Wierda WG, Kipps TJ, Khaw SL, et al. Substantial susceptibility of chronic lymphocytic leukemia to BCL2 inhibition: results of a phase I study of navitoclax in patients with relapsed or refractory disease. J Clin Oncol 2012;30:488–96.
- Babar IA, Cheng CJ, Booth CJ, Liang X, Weidhaas JB, Saltzman WM, et al. Nanoparticle-based therapy in an *in vivo* microRNA-155 (miR-155)-dependent mouse model of lymphoma. Proc Natl Acad Sci U S A 2012;109: E1695–704.
- Cheng CJ, Bahal R, Babar IA, Pincus Z, Barrera F, Liu C, et al. MicroRNA silencing for cancer therapy targeted to the tumour microenvironment. Nature 2015;518:107–10.
- Kong W, He L, Richards EJ, Challa S, Xu CX, Permuth-Wey J, et al. Upregulation of miRNA-155 promotes tumour angiogenesis by targeting VHL and is associated with poor prognosis and triple-negative breast cancer. Oncogene 2014;33:679–89.
- Levati L, Pagani E, Romani S, Castiglia D, Piccinni E, Covaciu C, et al. MicroRNA-155 targets the SKI gene in human melanoma cell lines. Pigment Cell Melanoma Res 2011;24:538–50.
- Li CL, Nie H, Wang M, Su LP, Li JF, Yu YY, et al. microRNA-155 is downregulated in gastric cancer cells and involved in cell metastasis. Oncol Rep 2012;27:1960–6.
- Qin W, Ren Q, Liu T, Huang Y, Wang J. MicroRNA-155 is a novel suppressor of ovarian cancer-initiating cells that targets CLDN1. FEBS Lett 2013;587: 1434–9.
- Palma CA, Al Sheikha D, Lim TK, Bryant A, Vu TT, Jayaswal V, et al. MicroRNA-155 as an inducer of apoptosis and cell differentiation in Acute Myeloid Leukaemia. Mol Cancer 2014;13:79.
- Chin LJ, Ratner E, Leng S, Zhai R, Nallur S, Babar I, et al. A SNP in a let-7 microRNA complementary site in the KRAS 3' untranslated region increases non-small cell lung cancer risk. Cancer Res 2008;68:8535–40.
- Saetrom P, Biesinger J, Li SM, Smith D, Thomas LF, Majzoub K, et al. A risk variant in an miR-125b binding site in BMPR1B is associated with breast cancer pathogenesis. Cancer Res 2009;69:7459–65.
- Ramsingh G, Koboldt DC, Trissal M, Chiappinelli KB, Wylie T, Koul S, et al. Complete characterization of the microRNAome in a patient with acute myeloid leukemia. Blood 2010;116:5316–26.
- Yu F, Jia X, Du F, Wang J, Wang Y, Ai W, et al. miR-155-deficient bone marrow promotes tumor metastasis. Mol Cancer Res 2013;11:923–36.
- 17. Zonari E, Pucci F, Saini M, Mazzieri R, Politi LS, Gentner B, et al. A role for miR-155 in enabling tumor-infiltrating innate immune cells to mount effective antitumor responses in mice. Blood 2013;122:243–52.
- Cubillos-Ruiz JR, Baird JR, Tesone AJ, Rutkowski MR, Scarlett UK, Camposeco-Jacobs AL, et al. Reprogramming tumor-associated dendritic cells in

Acknowledgments

The authors apologize to the many authors whose valuable contributions could not be cited due to space limitations.

Grant Support

Work in the Slack and Engelman labs is supported by grants from the NIH (R01-CA157749, R01-CA131301, and R01-GM073857). A.A. Svoronos is additionally supported by a NIH National Research Service Award (F30-CA196020) and Yale University's NIH Medical Scientist Training Program grant (T32-GM007205).

Received February 10, 2016; revised March 23, 2016; accepted April 4, 2016; published OnlineFirst June 20, 2016.

vivo using miRNA mimetics triggers protective immunity against ovarian cancer. Cancer Res 2012;72:1683–93.

- Kao CJ, Martiniez A, Shi XB, Yang J, Evans CP, Dobi A, et al. miR-30 as a tumor suppressor connects EGF/Src signal to ERG and EMT. Oncogene 2014;33:2495–503.
- Wu C, Jin B, Chen L, Zhuo D, Zhang Z, Gong K, et al. MiR-30d induces apoptosis and is regulated by the Akt/FOXO pathway in renal cell carcinoma. Cell Signal 2013;25:1212–21.
- 21. Gaziel-Sovran A, Segura MF, Di Micco R, Collins MK, Hanniford D, Vega-Saenz de Miera E, et al. miR-30b/30d regulation of GalNAc transferases enhances invasion and immunosuppression during metastasis. Cancer Cell 2011;20:104–18.
- Mitra AK, Zillhardt M, Hua Y, Tiwari P, Murmann AE, Peter ME, et al. MicroRNAs reprogram normal fibroblasts into cancer-associated fibroblasts in ovarian cancer. Cancer Discov 2012;2:1100–8.
- 23. Xu CX, Xu M, Tan L, Yang H, Permuth-Wey J, Kruk PA, et al. MicroRNA miR-214 regulates ovarian cancer cell stemness by targeting p53/Nanog. J Biol Chem 2012;287:34970–8.
- Yin Y, Cai X, Chen X, Liang H, Zhang Y, Li J, et al. Tumor-secreted miR-214 induces regulatory T cells: a major link between immune evasion and tumor growth. Cell Res 2014;24:1164–80.
- Xiang X, Zhuang X, Ju S, Zhang S, Jiang H, Mu J, et al. miR-155 promotes macroscopic tumor formation yet inhibits tumor dissemination from mammary fat pads to the lung by preventing EMT. Oncogene 2011;30: 3440–53.
- Tsai WC, Hsu PW, Lai TC, Chau GY, Lin CW, Chen CM, et al. MicroRNA-122, a tumor suppressor microRNA that regulates intrahepatic metastasis of hepatocellular carcinoma. Hepatology 2009;49: 1571–82.
- 27. Janssen HL, Reesink HW, Lawitz EJ, Zeuzem S, Rodriguez-Torres M, Patel K, et al. Treatment of HCV infection by targeting microRNA. N Engl J Med 2013;368:1685–94.
- Luna JM, Scheel TK, Danino T, Shaw KS, Mele A, Fak JJ, et al. Hepatitis C virus RNA functionally sequesters miR-122. Cell 2015;160:1099–110.
- 29. Gasparini P, Lovat F, Fassan M, Casadei L, Cascione L, Jacob NK, et al. Protective role of miR-155 in breast cancer through RAD51 targeting impairs homologous recombination after irradiation. Proc Natl Acad Sci U S A 2014;111:4536–41.
- Kutanzi KR, Yurchenko OV, Beland FA, Checkhun VF, Pogribny IP. Micro-RNA-mediated drug resistance in breast cancer. Clin Epigenetics 2011;2:171–85.
- Kalinowski FC, Brown RA, Ganda C, Giles KM, Epis MR, Horsham J, et al. microRNA-7: a tumor suppressor miRNA with therapeutic potential. Int J Biochem Cell Biol 2014;54:312–7.
- 32. Chou YT, Lin HH, Lien YC, Wang YH, Hong CF, Kao YR, et al. EGFR promotes lung tumorigenesis by activating miR-7 through a Ras/ERK/Myc pathway that targets the Ets2 transcriptional repressor ERF. Cancer Res 2010;70:8822–31.
- Pogribny IP, Filkowski JN, Tryndyak VP, Golubov A, Shpyleva SI, Kovalchuk O. Alterations of microRNAs and their targets are associated with acquired resistance of MCF-7 breast cancer cells to cisplatin. Int J Cancer 2010;127:1785–94.